

EFFECT OF ACTIVATED COCONUT SHELL CHARCOAL MEAL ON GROWTH PERFORMANCE AND NUTRIENT DIGESTIBILITY OF BROILER CHICKENS

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Abstract

A 56 days feeding trial was conducted on 225 Marshal Broiler chicks to evaluate the effect of activated coconut charcoal on their growth performance and nutrient digestibility. The birds were randomly allotted to five dietary treatments designated as T1, T2, T3, T4 and T5 containing 0, 0.5, 1, 1.5 and 2 % of activated coconut shell charcoal meal respectively. Each treatment had three replicate with 15 birds in each replicate. The birds were fed *ad libitum*. The result showed that feed intake and feed conversion ratio values significantly ($P<0.05$) different at the starter phase. The result of the finisher phase revealed significant ($P<0.05$) differences in the values of mean final body weight, body weight gain, feed conversion ratio and protein efficiency ratio. Also high percentage mortality was observed in birds fed 2 % activated coconut charcoal meal. The result of the nutrient digestibility showed no significant ($P>0.05$) difference in dry matter, crude protein, ether extract and nitrogen free extract. However, there were significant ($P<0.05$) differences observed in the digestibility of crude fibre and ash values. It was concluded that activated coconut shell charcoal supplementation at 2 % inclusion at the starter phase increases feed intake and final body weight. However, at the finisher phase activated coconut shell charcoal can be included up to 0.05 % inclusion level for better performance.

KEY words: Broilers, Activated coconut shell charcoal, Supplementation Performance

INTRODUCTION

More emphasis than ever is placed on global food security, both in terms of availability in the right nutrient proportions and affordability to the masses at a reasonable cost (Onibi *et al.*, 2009). Poultry meats offer considerable potential in bridging the gap between supply and demand for animal protein especially in developing countries like Nigeria. In this continent and other less developed nations, the low level of supply of animal protein is due to poor performance of livestock industry. (Onibi *et al.*, 2009) which has been attributed to factors such as inadequate

31 nutrition, high price and poor quality of feed industries. (Tewe.1997). However attempts have
32 been made to reduce the risk of mycotoxin in poultry feed which are secondary metabolites of
33 fungi, especially by species of *Aspergillus*, *Penicillium*, and *Alternaria*. Mycotoxins in feed can
34 cause huge problem for humans and animals, consumption of a mycotoxin-contaminated diet
35 may induce acute and long term chronic effects resulting in carcinogenic, estrogenic or immune
36 suppressive effects. Direct consequences of consumption of mycotoxin contaminated animal
37 feeds, include reduce feed intake, feed refusal, poor feed conversion, diminished body weight
38 gain, increase diseases incidence (due to immune suppression) and reduced reproductive capacity
39 (Matt Pearce *et al.*,2010) which leads to economic losses. The most common mycotoxins is
40 aflatoxins. The addition of mycotoxins binders to contaminated diets has been considered the
41 most promising dietary approach to reduce effects of mycotoxins (Afzal and Zahid, 2004)). The
42 theory is that the binder decontaminates mycotoxins in the feed by binding them strongly enough
43 to prevent toxic interaction with the consuming animals and prevent mycotoxin adsorption across
44 the digestive tract. It is impossible to completely remove such mycotoxins from the animal and
45 human food chain given that feed and its precursor (raw materials) can be stored and transported
46 through a range of time interval, atmosphere humidity and temperature, here lies the opportunity
47 for mycotoxins to do their detrimental work as well as during the cultivation and harvest stages
48 of grains.

49 Practical methods to detoxify mycotoxin contaminated grain in a larger scale and in a cost
50 effective manner is not currently available (Edrington *et al.*, 1997). A variety of physical,
51 chemical and biological techniques have been employed, however they have met with only
52 limited success. A recent approach has been the addition of non-nutritive adsorptive material to
53 the diet in order to reduce the absorption of mycotoxins from the gastro intestinal tract.

Activated charcoal is a form of carbon with an increase surface area for absorption. The term charcoal generally refers to carbonaceous residue of wood (Kutlu *et al.*, 2000). The very tiny, odorless, tasteless black powder is absorbent for many toxins, gases, drugs, and fat soluble substance without any specific action (Kutlu *et al.*, 2000). The adsorptive effect could be increased by treating with various substances at temperature ranging from 500-900° C, a treatment known as activation (Kutlu *et al.*, 2000). The final product is called activated charcoal (Osol,1975) as cited by (Ayanwale *et al.*, 2006). Charcoal acts as an insoluble carrier that non-specifically absorbed molecule, thereby preventing their absorption. Several researchers have tested the efficacy of activated charcoal in binding mycotoxins, there were many advantages of adding activated charcoal to animal diets, as it controls lactic acid concentration in the gastrointestinal tracts.(Ayanwale *et al.*,2006).

Materials and methods

A total of two hundred and twenty five (225) day old Marshal Chicks were used for the experiment. The birds were randomly allotted into five treatment groups of 45 birds and each group assigned to one of the dietary treatment of T₁ (0%), T₂ (0.5%), T₃ (1%), T₄ (1.5%) and T₅ (2%) activated coconut shell charcoal in a completely randomized design. Each treatment group was further subdivided into three replicate of 15 birds each. Feed and water was provided *ad libitum*, uniform light were provided 24 hours and proper vaccinations and medications was taken while body weight was recorded weekly. Feed conversion ratio, body weight gain were used as measures for birds growth performance. The experiment lasted for 8weeks. The birds were managed under a deep liter system with five (5) treatment groups and three replicates per treatment. 15 chicks were randomly allotted to each replicate in a complete randomized design arrangement. The housing System for the experiment was intensive. The birds were managed purely on

concrete floor with wood shavings on it, demarcated with wire mesh. The pen was thoroughly cleaned and disinfected before the commencement of the experiment. Heat was provided for the chicks during the brooding period with the use of charcoal pot and electric bulbs which were lit every evening and left all night. Vaccination and medication programme were strictly adhered to. Other Routine management procedures included, weighing of birds on arrival before the commencement of the experiment and subsequently at weekly intervals. Feed and clean water were supplied *ad-libitum*. Records of mortality, weight changes and feed intake were kept. The daily operation performed includes removal of left over feeds, cleaning of the drinkers and replacing the feeder and drinker with new feed and water. The following data were collected:

Data collection:

Data collected for ascertaining the performance of the birds includes; *initial body weight* which was collected by taking the weight of the day old birds at arrival in the farm using a weighing scale, *final body weight* was collected at the last day of the experiment using the same weighing scale, body weight gain was calculated by subtracting the initial body weight from the final body weight, feed intake was also calculated by subtracting the amount of feed refused from the amount of feed offered the birds and feed conversion ratio was calculated by dividing the feed intake by the body weight gain.

Digestibility trial

The digestibility trial was conducted by selecting two birds from each replicate of the treatments which were weighed and transferred into the metabolism cages. The birds continued to receive their diets for five days of the trial, the faecal droppings from each replicate separately collected daily were weighed and oven dried at 104^oc until a constant weight was obtained, After drying,

the droppings from each treatment were grouped together and sub-sample for proximate analysis (AOAC, 1990).

Statistical analysis: Data obtained from the experiment were subjected to the analysis of variance (ANOVA) in a completely randomized design using the procedure of SAS (2000).

Result and discussion

The experimental birds fed 2 % activated coconut shell charcoal supplementation had the superior growth performance when compare to those fed on other diets at the starter phase. However at the finisher phase, the superior growth was observed in birds fed 0.5% activated coconut shell charcoal supplementation. This variation in growth performance observed was consistent with the observation made by (Kutlu *et al.*, 2000) that activated charcoal supplementation at 2 % significantly increased body weight up to 28days of age, but at 49days of age activated charcoal inclusion at 2% did not significantly affect body weight.

The experimental birds showed no significant ($P>0.05$) difference in body weight gain at the starter phase, however at the finisher phase the significant ($P<0.05$) difference observed agrees with the report of (Wang, 2006). Whose report suggest that activated charcoal supplementation at 2% affected average weight gain not through feed intake, but possibly dilution of nutrient concentration or possible nutrient binding (Wang, 2006). The author reported that force feeding data with matured broiler roosters suggested less energy and protein availability when 2% activated charcoal was used in diet with less moldy corn.

The mean feed intake was significantly ($P<0.05$) different at the starter phase. However at the finisher phase feed intake was not significantly ($P>0.05$) different. This result is consistent with those reported by Baha Eddin (2007). Where there is considerable improvement in feed intake

up to 29 days of age and (Kutlu, *et al.*,2000) they also reported that oak charcoal showed similar result up to 28 days of age, this increase in feed intake might be the reason behind the increase in body weight at the starter phase.

Feed efficiency ratio of the birds fed the different diets showed that there was significant ($P<0.05$) difference observed in the feed conversion ratio with those fed activated coconut shell charcoal having higher ratio compared to the control diet both at the starter phase and finisher phase. The result is also in line with those reported by Kutlu *et al.* (2000) where oak charcoal showed similar result.

There was no significant ($P>0.05$) difference observed in protein efficiency ratio at the starter phase, however at the finisher phase there were significant ($P<0.05$) difference observed. The proteins were effectively utilized by birds fed 0.5 % activated coconut shell charcoal meal diet followed by 0 % inclusion of the activated coconut shell charcoal meal diet. This might be as result of possible nutrient binding or dilution of nutrient concentration. This result is consistent with the report of Wang, (2006) who stated that force feeding data with mature broiler rooster suggest less energy and protein availability when 2% activated charcoal was used in diet with less moldy corn. The percentage mortality was high on birds fed 2 % activated coconut shell charcoal meal diet at the finisher phase. While 0 % activated coconut shell charcoal supplementation and 0.5 % activated coconut shell charcoal supplementation had the least percentages of mortality. Also leg problem were observed in birds fed 1.5 % and 2 % activated coconut shell charcoal meal diet. These developments were also reported by Wang *et al.* (2006) who's report suggested that activated charcoal at 2% dosage could be harmful for the birds even in normal diets.

The significant ($P < 0.05$) difference observed in the digestibility of crude fibre and ash contents amongst birds fed activated coconut shell charcoal supplementation might be as a result of the crude fibre and ash content of the charcoal used in the diets which aids digestion in monogastric animals. Doll *et al.* (2004) documented that fibre has an influence on the development of the physical structure of the digestive system. Also Kutlu *et al.* (2000).reported that activated charcoal supplementation in broiler increases carcass ash content.

Conclusion

It is therefore concluded that broilers fed 2% activated coconut shell charcoal supplementation at the starter phase had beneficial effects on the body weight, feed intake, body weight gain and protein efficiency ratio, while at the finisher phase, although the birds all attained market weight of 2 kg weight and above, birds fed 0.5 % activated coconut shell charcoal supplementation had better performance amongst the parameters measured.

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Table 1 Composition of the basal diets.

Ingredients	Starter phase (%)	Finisher phase (%)
Maize	51.63	59.67
Soybean meal	18.82	14.27
GNC	17.21	13.10
Fish	4.00	3.00
Wheat offal	2.62	5.00
Bone meal	2.00	2.00
Oyster shell	1.00	1.00
Premix	0.30	0.30
Methionine	0.20	0.20
Lysine	0.20	0.20
Salt	0.30	0.30
Palm oil	2.00	1.00
Total	100	100

Calculated values

Crude protein 24.0 1%,

Metabolizable energy 3150 (kcal kg^{-1})

Each 2.5kg contain vita. A – 10,000,000iu.; vita. D-2,000,000iv, vita. E- 20,000iv, vita. k- 2,250mg; thiamine –

17omg; 12iboflavin – 5,000mg; Pyridoxine – 2,750mg; Niacin- 27,500mg; vit B12 -15mg; Pantothenic acid –

7,500mg; folic Acid – 7,500mg; Biotin – 50mg; manganese – 80g; zine – 50g; copper – 5g; iodine 1.5g; selenium –

200mg and cobalt – 200mg;

Activated coconut shell charcoal supplementation (starter and finisher phase) at**0 %, 0.5 %, 1 %, 1.5 %, and 2 % per 100kg of diet.**

Table 4.4 Growth Performance of broilers fed diets supplemented with Activated coconut shell charcoal. (Starter phase)

Parameters	T ₁	T ₂	T ₃	T ₄	T ₅	SEM
Initial body weight(g)	33.33	33.33	33.33	33.33	33.33	0.00
Final body weight(g)	889.88	880.78	834.35	880.37	903.97	12.66
Mean body weight gain(g)	218.61	211.95	200.29	199.60	209.43	12.53
Mean feed intake(g)	387.66 ^b	401.28 ^b	381.62 ^b	382.68 ^b	445.82 ^a	7.15
Feed conversion ratio	1.76 ^b	1.82 ^{ab}	1.82 ^{ab}	1.84 ^{ab}	1.92 ^a	0.20
Protein efficiency ratio	7.34	8.02	7.99	8.17	8.34	0.14

Means on the same row with different superscript are significantly different (P<0.05)

T₁ –0% activated charcoal supplementation. T₂-0.5% activated charcoal supplementation.T₃ -1% activated charcoal supplementation. T₄-1.5% activated charcoal supplementation.T₅ .2% activated charcoal supplementation.

Table 4.5.Growth Performance of broilers fed diets supplemented with Activated coconut shell charcoal. (Finisher phase)

Parameters	T ₁	T ₂	T ₃	T ₄	T ₅	SEM
Initial body weight(g)	889.88	880.78	834.45	830.37	904.07	12.65
Final body weight(g)	2324.70 ^a	2378.50 ^a	2284.90 ^a	2131.40 ^a	2034.60 ^b	38.39
Mean body weight gain(g)	358.59 ^{ab}	382.74 ^a	362.59 ^{ab}	325.29 ^{ab}	282.60 ^b	13.20
Mean feed intake(g)	964.04	967.39	868.91	899.65	915.52	15.79
Feed conversion ratio.	2.92 ^b	2.32 ^c	2.74 ^{ab}	2.96 ^b	3.75 ^a	0.01
Protein efficiency ratio.	7.53 ^{ab}	8.11 ^a	7.45 ^{ab}	7.01 ^{ab}	6.57 ^b	0.16
Mortality (%).	7.00	7.00	8.90	10.55	12.22	

Means on the same row with different superscript are significantly different (P<0.05)

T₁-0% activated charcoal supplementation. T₂-0.5% activated charcoal supplementation.

T₃-1% activated charcoal supplementation. T₄-1.5% activated charcoal supplementation.

T₅-2% activated charcoal supplementation.

Table 4.6 Nutrient digestibility of broilers fed diets supplemented with Activated coconut shell charcoal

Parameters	T ₁	T ₂	T ₃	T ₄	T ₅	SEM
Dry matter	81.46	83.07	81.87	79.78	78.84	0.84
Crude protein	92.21	93.50	91.25	89.96	88.62	0.88
Crude fibre	77.2 ^{ab}	72.86 ^b	85.91 ^a	89.96 ^a	88.62 ^a	2.33
Ether extract	90.37	90.02	91.35	92.42	91.11	0.71
Ash	77.32 ^{ab}	71.88 ^b	78.92 ^{ab}	80.23 ^{ab}	81.23 ^a	1.33
NFE	70.40	76.03	75.19	72.05	71.90	0.81

Means on the same row with different superscript are significantly different (P<0.05)

T₁-0% activated charcoal supplementation. T₂-0.5% activated charcoal supplementation.

T₃-1% activated charcoal supplementation. T₄-1.5% activated charcoal supplementation.

T₅-2% activated charcoal supplementation.

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